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Cover cropping in integrated weed management

Dissertation Submitted in fulfillment of the requirements for the degree "Doktor der Agrarwissenschaften" (Dr. sc. agr. / Ph.D. in Agricultural Sciences) to the Faculty of Agricultural Sciences

by

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Für meine Eltern

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Chapter I

General Introduction

1 General Introduction

The integrated weed management implements the careful selection of appropriate crop protection methods to counteract the emergence and spread of weed populations within the field [1]. This management system aims to provide healthy, high-yielding crops by combining biological, mechanical and chemical approaches with minimal disturbance of the agricultural ecosystem [2]. The integration of cover crops and living mulches into crop rotation can be useful for the ecologic and economic production of food, while providing various ecological services [3]. Cover cropping between two main crops contributes to the agricultural production by reducing soil erosion [4] and nitrate leaching [5,6], improving soil fertility and structure [7,8], enhancing the microbial activity [9] and biological weed suppression [4,10–13]. Studies reported weed suppression by more than 90% for winter annual weeds and 40% for perennial weeds during autumn and winter after cover crop cultivation [12,14–19]. Cover crop and living mulch inclusion in the crop rotation provides three opportunities to interfere with the lifecycle of weeds [20].

The first result of cover crop inclusion is the inhibition of weed germination, growth and seed production in autumn as a result of the competition of cover crops and weeds for limited resources as light, water, space and nutrients and allelopathic effects [19]. Allelopathy is described as any process involving secondary metabolites, produced by plants, which were released in the environment and influence the development and growth of adjacent plants in agricultural systems [21]. These substances can be introduced actively during the cover crop germination and growth via various pathways as root exudation, volatilization or by the leaching from plant biomass [22,23]. Allelochemicals are diverse in their chemical structure and were identified and isolated in many cover crops [23–25]. For example, members of the family of Poaceae, like Avena sp. and Secale sp., were reported to exude phytochemicals with high allelopathic effects on weeds [26-29]. Schulz et al. identified 16 allelochemicals in rye (Secale cereale L.), including the benzoxazinones, and emphasized their high allelopathic contribution to the overall weed suppression [28]. The complete pathway and dynamics from the exudates of the donor plant, the transformation, and degradation processes to the uptake by the target plant is extremely complex and requires intensive research [27].

The allelopathy phenomenon was investigated in terms of inducibility by biotic and abiotic factors [29]. Pathogen or insect attacks were studied as elicitor for biotic-induced and -enhanced allelopathy in conjunction with forced release rates of allelochemicals or higher gene expression that regulates the biosynthesis of the phytotoxic compounds [27,29-31]. Beside the biotic factors, changes in the environment as temperature, humidity, nutrients, mechanical damage or irradiation can also increase the accumulation and expression of allelochemicals [20,29,32,33]. Among other weed suppressing effects, the clear understanding of the proportions of allelopathic effects on the overall weed suppression under natural conditions is lacking [34]. The knowledge about the contribution of allelopathic and competitive effects of different cover crops could enable farmers to suppress specific weed species or communities with appropriate cover crop mixtures in autumn and winter. Additionally, a mixture of different cover crop species is more flexible to unpredicted biotic and abiotic stressors due to a higher elasticity and ability to recovery compared to a mono cultivation. From this might also follow a more effective weed suppression [35]. Furthermore, cropping methods, like an optimum cover crop sowing date and fertilization, may enhance the beneficial effects in agricultural systems.

In spring, the cover crops froze or were sprayed with non-selective herbicides to induce the formation of mulch which provides the second opportunity for interference in the weeds lifecycle. The cover crop residues on the soil surface offer ecological benefits like reducing the soil evaporation and erosion [36–39], decreasing daily soil temperature excursion [40–42] and suppressing weeds [42–45]. Cover crop mulch alters physically the weed seeds environment by changes in light availability, humidity, nutrient mobilization, soil temperature, soil moisture and can also offer additional allelopathic effects [17]. Especially cover crops with high allelopathic properties seem to be well-suited for suppression of weed germination and growth [46]. Moreover, the incorporation and the associated mechanical wounding of these cover crops could enhance the biochemical weed suppression in spring [20]. Therefore, a combination of diverse cover crops with optimum physical and allelopathic traits could provide higher

weed control efficacy due to a combination of different inhibitory mechanisms prior to the main crop sowing.

The biological interference with weed development by cover crops from autumn to spring can be continued by the integration of living mulches in the main crop in summer, which provides the third opportunity. Living mulches are cover crops, which are sown simultaneously with or shortly after sowing the main crop [4,47]. The level of competition for natural resources between the undersown cover crop and the main crop depends on the cover crop species and must be carefully selected to avoid quantitative and qualitative yield losses [48]. Especially in main crops with wide row distances, as sugar beets, living mulches provide similar ecological benefits compared to summer or autumn sown cover crops including the inhibition of weed germination and development [4,47–52]. Furthermore, the substantial weed suppression during the growth of the main crop could reduce herbicide input which contributes to environmentally sustainable agriculture.

1.1 Objectives

In the presented thesis, the main objectives were

- to investigate the weed suppressing effects of different cover crops under mono and mixture cultivation in autumn and winter
- to optimize cover crop weed control by different cover crop sowing dates and fertilization
- to explore the impact of diverse cover crop mulches on sugar beet development and the germination and growth of specific weed species
- to test the feasibility of the cultivation of living mulches in sugar beet for weed suppression with respect to sugar beet quality and quantity parameters
- to evaluate the contribution of competitive and biochemical effects on the overall weed suppression by cover crops
- to identify susceptibilities of different weed species to specific cover crops

1.2 Structure of the dissertation

The current thesis consists of ten chapters contributing to the optimization and exploration of approaches for integrated weed management with cover crops. The thesis begins with the general introduction (Chapter I) presenting the field of research and emphasizing the objectives of this work. The following chapters (Chapters II-VII) are six research articles composing the main work of this thesis.

The scientific articles were arranged along an agricultural vegetation period from late summer over spring to the following summer and present the opportunities and strategies for interference in the lifecycle of weeds. In Chapter II and III, cover crops were investigated on their weed suppressive ability in autumn and winter concerning the cultivation in mono and mixture cultivation as well as the impact of cover crop sowing date and fertilization. Chapter IV and V deal with the role of competitive and biochemical effects on the overall weed suppression and demonstrates the sensitivity of specific weeds to biochemical stresses. Chapter VI presents the potential of different cover crop mulches to suppress weeds in the early development of sugar beet crops. In Chapter VII, the possibility of living mulch cultivation in sugar beet crops was tested with respect to weed suppression and quantity and quality parameters of the main crop. The general discussion (Chapter VIII) gives a critical overview of the research articles. The whole thesis is summarized in Chapter IX.

Apart from the peer-reviewed journal articles, three more contributions to national and international scientific conferences were presented as an oral presentation during the course of this thesis. This work was supplementary to the included articles and therefore not included in the current thesis.

- Sturm, D.J. & Gerhards, R. (2016). Comparison of different cover crop mulches and extracts on inhibition of crop and weed growth. In: *Proceedings of the 27th German Conference of Weed Biology and Weed Control*, 452, 424-430.
- Sturm, D. J., Kunz, C., & Gerhards, R. (2016). Comparison of different cultivations of *R. sativus* var. *oleiformis* as cover crop on weed

suppression. In: *Proceedings of the* 7th *International Weed Science Congress*.

• Kunz, C., Sturm, D. J., & Gerhards, R. (2016). Effect of Strip Tillage Systems on weed suppression in sugar beets by utilizing different cover crops. In: *Proceedings of the 7th International Weed Science Congress*.

Chapter II

Do cover crop sowing date and fertilization affect field weed suppression?

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2 Do cover crop sowing date and fertilization affect field weed suppression?

Summary

The weed suppressive ability of oilseed radish (Raphanus sativus var. oleiformis Pers.) cover crop is attributed to high competitiveness for resources and allelopathic effects on weeds. The oilseed radish cover crop was sown in five treatments plus an untreated control over a period of five weeks before and three weeks after winter wheat harvest. Additionally, fertilization effects on oilseed radish biomass and weed suppression were measured. The highest biomass of the cover crop was observed 12 weeks after harvest (WAH) when the oilseed radish was sown one week after harvest (1 WAH) (2015) and five weeks before harvest (5 WBH) (2016). No differences of fertilization were observed concerning oilseed radish and weed biomass in 2015, whereby increased biomass was found after fertilization in 2016. The highest weed control efficacy of up to 83% and 90% was achieved in treatments 1 WAH (2015) and 5 WBH (2016) at 12 WAH. The early sowing of oilseed radish in winter wheat resulted in low germination and biomass yield within the field, due to low precipitation in 2015. Nevertheless, there is a high potential of early sown oilseed radish for higher weed control efficacy, which was demonstrated in 2016.

Keywords: allelopathy, Brassicaceae, intercropping, cropping system, competition, weed density

Chapter III

Allelopathic effects and weed suppressive ability of cover crops

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3 Allelopathic effects and weed suppressive ability of cover crops

Summary

Field and laboratory experiments were conducted to investigate the weed suppressing effects of cover crops in single and mixed cultivation. Weed densities in the field experiments ranged from 0 to 267 plants m⁻² with *Chenopodium album* L., Matricaria chamomilla L., Stellaria media (L.) Vill. as predominant weeds. It was found that mustard (Sinapis alba L.), fodder radish (Raphanus sativus var. niger J. Kern) and spring vetch (Vicia sativa L.) suppressed weeds by 60% and cover crop mixtures controlled weeds by 66% during the fallow period at three experimental locations in 2013, 2014 and 2015. The allelopathic effect of the same cover crops/mixtures on weed growth was analyzed in laboratory experiments. Aqueous cover crop extracts were applied on weeds and analyzed using LC/MS/MS. Mean germination time, germination rate and root length of weeds were determined. Extracts prolonged the germination time by 54% compared to the control with only water. In all cases, inhibitory effects on germination rate and root length were measured. Weed density in the field was found to be correlated with the root length in the germination tests. Our work reveals that biochemical effects play a major role in weed suppression of cover crops.

Keywords: allelopathy, erosion, root growth, competition, intercropping

Chapter IV

Contribution of competitive and biochemical effects by different cover crops to weed suppression

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4 Contribution of competitive and biochemical effects by different cover crops to weed suppression

Summary

Cover crops can suppress weeds within agricultural fields due to competitive and biochemical effects. Greenhouse experiments were conducted to evaluate the relative proportions of both effects to the total weed inhibition. Six different cover crop species were combined with three weed species in the presence or absence of active carbon over a period of four weeks. Active carbon was used as an adsorbent for biochemical substances in the soil. Our study reveals that the balance of competition between cover crops and weeds shifted when biochemical effects in the soil were minimized by active carbon. We assume that the degree of cover crops biochemical effects on weeds is species-specific, both on the side of cover crops, as well as on the weed side. The knowledge about the contribution of competitive and biochemical effects by cover crops would enable us to create cover crop mixtures to suppress specific weed species and communities.

Keywords: Allelopathy, *Alopecurus myosuroides*, Intercropping, *Stellaria media*, *Triticum aestivum*, Weed control

4.2 Introduction

Cover cropping provides ecological and economic benefits in agricultural fields, including nutrient recycling, reduction of soil erosion and effective weed suppression with a potential contribution to Integrated Weed Management [4]. The weed suppressive ability of cover crops is characterized by a high competition for light, water, space and nutrients attributed to a combination of competitive and biochemical effects [11]. Several studies have shown that allelopathy can play an important role in the overall weed suppression and the competition for limited plant resources [11]. Allelopathy is the chemically mediated interference between co-occurring plants and includes a growth stimulation or inhibition of the target-plant, mostly following a hormesis [53]. Callaway & Ashehoug reported stronger allelopathic effects of the invasive plant Centaurea diffusa Lam. on different grass species in North America compared to grass species to which C. diffusa is native [54]. Based on that concept, cover crops are non-coevolved competitors to weeds, therefore weeds lack a natural adaption to their novel phytochemicals. Therefore, greater allelopathic interference can be expected when an allelopathic plant occurs in a non-native range [55]. Some cover crop species were investigated for the active release of allelopathic compounds as the family of Poaceae [28], Fagopyrum sp. [56] or Helianthus annuus L. [57]. The general effects of competition within the field are partly well understood as isolated mechanisms, but there is a lack of information about the relative proportions on the total weed suppressive effects [58].

The aim of this study was to estimate the relative proportions of biochemical and competitive effects of six different cover crops on the overall weed suppression of two weed species (*Stellaria media* (L.) Vill. and *Alopecurus myosuroides* Huds.) and volunteer wheat (*Triticum aestivum* L.) in greenhouse trials. In order to determine the allelopathic capability of the selected cover crops, the weeds were grown in the presence and absence of the potential allelopathic competitor (with and without cover crops) in the presence or absence of active carbon, as an adsorbent for biochemical compounds, in the soil. The knowledge about the proportions of biochemical and competitive effects could enable us to create cover

crop mixtures with optimum traits for more effective specific weed suppression within the field.

4.3 Material and Methods

4.3.1 Experimental set-up

The greenhouse experiments were carried out at the University of Hohenheim from 2015-2016 to estimate the relative contribution of competitive effects and biochemical traits to the overall weed suppression on different weed species (S. media and A. myosuroides) and volunteer wheat (T. aestivum). A soil with 60% sand and 40% turf (v/v) was prepared (Soil-N). Pulverized active carbon (Carl Roth GmbH + Co. KG, Karlsruhe, Germany), with a particle size of < 0.8 mm, was mixed with half of the Soil-N to reach a concentration of 6% (v/v) (Soil-AC). The addition of active carbon was performed to reduce potential biochemical interference in the soil between cover crops and weeds [54]. Active carbon is used as adsorbent due to the high porosity and adsorptive capacity for many organic compounds. An enhanced weed growth after soil implementation with active carbon indicates the presence of allelopathic compounds released by the cover crops [55]. Both soils were filled in 2-L pots separately and one gram of a slow release fertilizer (16:9:12 N:P:K, Osmocote®, Scotts Celaflor GmbH, Mainz, Germany) was added to each pot. This was performed to reduce possible effects of the active carbon on the nutrient availability in the soil [59]. The different cover crops (Tab. 7) were sown separately in pots, using recommended sowing rates, together with 40 seeds of A. myosuroides or 30 seeds of S. media (Herbiseed, Reading, UK) or 7 seeds of winter wheat (T. aestivum cv. 'Pamier'). Shortly after germination, weeds were thinned out to ten and the winter wheat to five plants per pot. Two controls with Soil-N (Control-N) and Soil-AC (Control-AC), plus weeds and no cover crops were prepared to determine confounding effects of the active carbon on weed growth. The greenhouse setup was 12 h / 12 h (day/night) with the temperatures being respectively 20/15 °C. All pots were irrigated daily with tap water to field capacity. The pots were arranged in a randomized complete block design with four replicates and the experiment was

repeated in time. Cover crop and weed biomass were harvested after four weeks and were dried at 80°C for 48 h before being weighed.

Cover crop	Scientific name	Plant Family	Sowing rate [kg ha ⁻¹]
Oilseed radish	Raphanus sativus var. oleiformis Pers.	Brassicaceae	20
Buckwheat	Fagopyrum esculentum Moench	Polygonaceae	45
Black Oat	Avena strigosa Schreb.	Poaceae	100
Common flax	Linum usitatissimum L.	Linaceae	100
Ramtil	Guizotia abyssinica (L.f.) Cass.	Asteraceae	8
Sunflower	Helianthus annuus L.	Asteraceae	25

 Table 1 Different cover crop treatments and sowing rates in the greenhouse experiments.

4.3.2 Statistical Analysis

The data of the greenhouse experiments were analyzed with the statistical language R version 3.1.1 [60] with a linear mixed effects model taking soil type and cover crop as different factors. The homogeneity of variance and the normal distribution were checked visually and transformations of data were performed, if necessary. An analysis of variance (ANOVA) was carried out to test the effects of active carbon, cover crop species and their interaction. Years and replications were considered random effects. Further, separate analyses were performed for each weed species. Means were separated via a Tukey's *HSD* test at $P \le 0.05$. The proportions of competitive and biochemical effects in the greenhouse experiments were calculated as following:

Overall weed suppression [%] =

1 - $\frac{\text{Weed biomass with cover crop}}{\text{Weed biomass without cover crop}} * 100$

Biochemical weed suppression [%] =

Overall weed suppression in Soil-N [%] - Overall weed suppression in Soil-AC [%]

Biochemical effects are considered significant, if significant differences between weed biomass in Soil-N and Soil-AC and no statistical differences between Control-N and Control-AC were observed.

4.4 Results

4.4.1 Weed suppressive effects of the different cover crops

The utilization of active carbon as an adsorbent for organic compounds to minimize biochemical effects in the soil represents a suitable approach for the evaluation of biochemical effects on the overall weed suppression.

The weed suppressive abilities of the selected cover crops were tested in the prepared soil (Soil-N) and the soil supplemented with 6% of active carbon (Soil-AC). The three weeds *A. myosuroides*, *S. media* and *T. aestivum* were suppressed by the different cover crops (Fig. 6). In all cases, Control-N showed no differences to Control-AC. The biomass of all weeds was significantly reduced in all cover crop treatments compared to the controls.

S. media biomass was reduced by 21 to 80% with the most effective growth suppression caused by *H. annuus* (80%), *F. esculentum* (77%) and *R. sativus* (59%) across both soil treatments. Suppression of *S. media* by the cover crops *R. sativus*, *F. esculentum*, *A. strigosa* and *H. annuus* was significantly lower in Soil-AC. No suppression was found for *L. usitatissimum* and *G. abyssinica*. The biomass of the weed *A. myosuroides* was significantly reduced among all cover crops with up to 89% by treatment *R. sativus*. Significant differences between both different soils were observed for treatment *F. esculentum*, only. The average weed control efficacy of *T. aestivum* was 54 and 48% in Soil-N and Soil-AC across all cover crop treatments. No statistical differences between Soil-N and Soil-AC were observed, except for *A. strigosa* which reduced the growth of *T. aestivum* by 67 (Soil-N) and 48% (Soil-AC).



Figure 1 Overall plant biomass reduction [%] of the weeds *S. media*, *A. myosuroides* and *T. aestivum* cultivated in untreated soil (Soil-N) and soil containing 6% active carbon (Soil-AC) with different cover crops after a period of four weeks in greenhouse trials. Means with identical letters within the graph do not differ significantly based on the Tukey HSD test (p < 0.05).

The biomass of all cover crops growing in the presence of the weeds was not affected by both soils (Fig. 7). The highest biomass was achieved by *H. annuus* and *F. esculentum* with meanly 4.0 and 3.5 g pot⁻¹ across all weeds.



Figure 2 Biomass [g pot⁻¹] of the different cover crops cultivated in untreated soil (Soil-N) and soil containing 6% active carbon (Soil-AC) with different weeds after a period of four weeks in greenhouse trials. Means with identical letters within the graph do not differ significantly based on the Tukey HSD test (p < 0.05).

4.4.2 Proportions of biochemical effects on the overall weed suppression

The differences between the overall weed suppression (Soil-N) and the competitive weed suppression (Soil-AC) provides an estimation of the proportion of biochemical effects on the total interference between specific cover crop and weed species (Tab. 8).

Table 2 Proportions of biochemical effects [%] on the overall weed suppression by different cover crops in greenhouse trials (experiment 2). The calculated biochemical effects were only significant [*] if significant differences between the weed biomass in both soils (Soil-N and Soil-AC) and no differences between both controls were detected.

Cover crop	Biochemical	Biochemical weed suppression [%]	
	S. media	A. myosuroides	T. aestivum
R. sativus var.	28.1 *	2.5	1.1
oleiformis			
F. esculentum	13.7 *	12.2 *	1.3
A. strigosa	25.5 *	4.4	19.2 *
L. usitatissimum	0	5.5	2.0
G. abyssinica	1.4	8.1	5.5
H. annuus	11.5 *	6.2	7.6

Significant biochemical effects by 11.5, 13.0, 22.4 and 28.1% were calculated for *H. annuus*, *F. esculentum*, *A. strigosa* and *R. sativus* cover crops, respectively. The weed *S. media* was found to represent the most sensitive weed species to biochemical stress. The cover crops *F. esculentum* and *A. strigosa* showed significant proportions of biochemical effects on the overall weed suppression of *A. myosuroides* and *T. aestivum*, only. The two cover crops *L. usitatissimum* and *G. abyssinica* showed no significant biochemical effects on the weeds in this experiment.

4.5 Discussion

In this study, the biomass of some weeds growing with different cover crop treatments was significantly increased when active carbon was present in the soil. Correspondingly, the active carbon gave some cover crops a competitive disadvantage against the weeds. The observed effects of active carbon on the cover crops weed suppression can be interpreted as an evidence for allelopathy [54]. However, weed growth influencing effects by soil microbiota due to the addition of active carbon may be possible [54].

Some of the tested cover crops showed were known for the active release of allelochemicals during growth. Studies about the weed suppressive ability of F. esculentum within the field reported of reduced weed biomass with differences between various weed species [56]. A common explanation for the weed inhibiting effects is the competition for light and nutrients [61,62]. However, no experiments could prove this hypothesis, so far [56] and a significant competition for nutrients, as a factor for observed weed suppression, can be neglected in this experiment due to fertilization. This was shown in field experiments with high soil nutrient supplies resulted in an effective weed control by F. esculentum [62]. Kalinova et al. demonstrated that the high weed suppressive ability of F. esculentum from germination to early development originates from the root exudation of several phytotoxic substances [63]. Falquet et al. showed that light competition and root interaction between F. esculentum and the weed (Amaranthus retroflexus L.) led to significant weed biomass reductions, while excluding other suppressive factors as the competition for nutrients and water [64]. Further, the authors concluded the major role of the root interaction of F. esculentum, including the release of allelopathic root exudates, on the overall weed suppression compared to the light competition.

The family of Poaceae, including *A. strigosa*, has been documented to release allelopathic substances in the rhizosphere [28]. Substances of the chemical groups of benzoxazinones and several phenolic acids were actively exudated via the roots during growth and were tested for inhibitory effects on weed growth [28]. *H. annuus* has been investigated extensively for inherent allelopathic substances in various plant parts which might be able to influence the germination and growth of weeds [57]. Glucosinolates and their degradation products are the main sources

for allelopathic effects on weeds by *R. sativus*. Several studies reported of high amounts of glucosinolate degradation products after tissue damage or the incorporation of *R. sativus* residues in the soil [65]. Alternatively, the active exudation of inhibitory substances via the roots or leachates from the leaves could be possible, as shown by the exudation of 2-phenylethyl isothiocyanate and allyl isothiocyanate by *Brassica napus* L. roots [65]. The cover crops *L. usitatissimum* and *G. abyssinica* showed a strong weed suppressive ability with no significant differences in the weed suppression in Soil-N and Soil-AC. This can be attributed to the absence of root or shoot exudation of inhibitory compounds in the soil. Moreover, there is a lack of information about potential allelochemicals and their exudation by *L. usitatissimum* and *G. abyssinica*, so far. Therefore, these two cover crop species exert weed suppression mainly by physical competition. However, there is no evidence for the volatilization of allelopathic compounds during growth induced by all investigated cover crops. Therefore, the mentioned release pathway can be neglected in this experiment.

In this experiment, *A. strigosa* showed significant substantial biochemical effects on the weeds *S. media* and *T. aestivum*. The absence of these effects on *A. myosuroides* may be attributed to a higher detoxification activity of this weed against growth suppressing allelochemicals like benzoxazolin-2(3H)-one (BOA) released by *A. strigosa* [28]. The accumulation of multi drug resistance transporters as verapamil, nifedipine and ethacrynic acid with glutathione transferases in a donor plant may enhance the extrusion of phytotoxic compounds, as BOA, out of the protoplasts [28]. In general, allelopathic effects were speciesspecific [11,28,66]. In this experiment, the weed *S. media* showed the greatest sensitivity to biochemical effects. Earlier studies confirm the increased biochemical susceptibility of *S. media* compared to other weeds [66]. This is also in line with other studies, which demonstrated a higher sensitivity of dicotyledonous weeds against the presence of *F. esculentum* seedlings and its released allelochemicals [63].

The proportions of competitive and biochemical effects on the overall weed suppression indicates an important role of the biochemical effects. Nevertheless, the competitive effects showed a higher contribution to the overall effects. Therefore, a rapid cover crop germination and development combined with a dense canopy and high soil coverage is a prerequisite for an effective weed suppression within the field.

Future research should investigate if competitive and biochemical effects provide additive or synergistic effects on the overall weed suppression. Moreover, further cover crop species and cultivars should be investigated with strong biochemical effects on different weeds.

Chapter V

Inhibitory effects of cover crop mulch on germination and growth of *Stellaria media* (L.) Vill., *Chenopodium album* L. and *Matricaria chamomilla* L.

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The original publication is available at: https://www.journals.elsevier.com/crop-protection/ doi: 10.1016/j.cropro.2016.08.032 5 Inhibitory effects of cover crop mulch on germination and growth of *Stellaria media* (L.) Vill., *Chenopodium album* L. and *Matricaria chamomilla* L.

Summary

Cover crops may suppress weeds due to their competitive effects and the release of inhibitory compounds. We examined the inhibitory influence of 11 cover crop mulches on the germination and growth of weed species (Stellaria media (L.) Vill., Chenopodium album L. and Matricaria chamomilla L.) in laboratory, greenhouse and field experiments. In the laboratory, cover crop extracts were tested in germination bioassays at six concentrations (0 to 500 mg ml⁻¹). The germination rate and root length (i) were measured 10 days after treatment (DAT). Pot experiments were carried out in the greenhouse to investigate the effects of cover crop mulch (ii) incorporated into the soil on weed germination and weed dry mass. Field trials measured the weed suppressive effects of cover crops and cover crop mixtures (iii). Correlations were determined between the experiments to quantify the competition and the biochemical effects of cover crops separately. Cover crop extracts at a concentration of 125 mg ml^{-1} (i) significantly reduced the weed germination rate by 47% and the root length by 32% on average. M. chamomilla showed a lower susceptibility to the extracts of S. alba, R. sativus var. niger and H. annuus compared to C. album and S. media. The mulch-soil mixtures (ii) significantly reduced the germination rate by 50% and the dry mass by 47% on average across all three weed species, while *M. chamomilla* showed the highest tolerance to the mulches of V. sativa and A. strigosa. The correlation analysis revealed a strong positive correlation between extract toxicity and field weed suppression and, thus, indicated a high impact of the allelopathic effects of the tested cover crops on weed suppression, especially for S. media and M. chamomilla.

Keywords: allelopathy, germination test, phytotoxicity, plant extracts, root length, sugar beet

Chapter VI

Weed suppression and early sugar beet development under different cover crop mulches

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Summary

Field experiments were conducted at two locations in 2014-2015 and 2015-2016 to investigate the weed suppressive ability of cover crop mulches in sugar beets. Three cover crops and two cover crop mixtures were tested in all four field experiments. The weed densities ranged from 2 up to 210 plants m⁻² with *Chenopodium album* L. and *Stellaria media* (L.) Vill. as predominant species. *Sinapis alba* grew significantly faster than *Vicia sativa*, *Raphanus sativus* var. *niger* and both cover crop mixtures. *Sinapis alba*, *Vicia sativa*, *Raphanus sativus* var. *niger* reduced weed density by 57, 22 and 15% across all locations. The mixture of seven different cover crops observed a reduced weed emergence of 64% compared to the control plot without cover crop mulch. The early sugar beet growth was enhanced by all mulch treatments in 2015 and decelerated in 2016.

Keywords: *Beta vulgaris, Chenopodium album,* conservation tillage, cover crop mixture, integrated weed management, intercropping, *Stellaria media*

Chapter VII

Weed Suppression of Living Mulch in Sugar Beets

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7 Weed Suppression of Living Mulch in Sugar Beets

Summary

Weed suppression in sugar beets (Beta vulgaris.) is commonly achieved with two to three post-emergent herbicide applications across the entire field. Field studies were performed in order to investigate the weed suppressing ability of *Medicago* lupulina, Trifolium subterraneum and a mixture of Lolium perenne and Festuca pratensis as living mulches in sugar beet at four locations in southern Germany during 2014 and 2015. Living mulches were sown 2 and 30 days after sowing (DAS) of sugar beets. Weed densities ranged from 0 to 143 plants m^{-2} with Chenopodium album, Polygonum convolvulus and Polygonum aviculare being the most abundant weed species. It has been found that living mulches could reduce herbicide input up to 65%. Weed suppression of living mulch was highest with Trifolium subterraneum (71%). The early sown living mulches (2 DAS) revealed a 28 g m⁻² higher biomass compared to late sowing (30 DAS). However, no any linear correlation was found between living mulch biomass and weed suppression. The white sugar yield (WSY) was highest in the herbicide treatments (12.6 t ha⁻¹). Trifolium subterraneum yielded the highest WSY of the living mulches with 11.1 t ha⁻¹ across all locations. Our work reveals that living mulch can play a major role in integrated weed management by reducing herbicides in sugar beet production.

Keywords Biomass, *Beta vulgaris*, Cover crop, *Festuca*, *Lolium*, *Trifolium*, Intercropping, Sugar content, Sugar yield, Weed density

Chapter VIII

General Discussion

8 General Discussion

The integration of cover crops and living mulches in the crop rotation can provide a substantial biological inhibition of weed germination and growth. The aim of this work was the optimization and exploration of integrated weed control strategies with cover crops. For this purpose, three laboratory, two greenhouse and five field experiments were performed from 2014-2017 at the University of Hohenheim, Germany.

We examined the interference in the weed lifecycles by the cultivation of different cover crops from autumn to the following summer. Furthermore, attention is directed to the contribution of altering the cropping system, including sowing date and fertilization. Laboratory and greenhouse experiments provided a closer examination of the proportions of biochemical effects of cover crops on the overall weed suppression which will be discussed with regard to possible benefits for farmers.

8.1 Weed suppression by cover crops in autumn and winter

In the experiments, cover crops were able to suppress weed density and biomass by up to 91 and 89% in autumn and winter due to a combination of competitive and biochemical effects. In chapters 2 and 3, the investigated cover crops showed varying inhibitory effects on weed density and biomass depending on mono or mixture cultivation, cover crop species and sowing date. The mono cultivation of the cover crops *S. alba*, *R. sativus* var. *niger* and *V. sativa* resulted in a weed density reduction of 60%, while a weed suppression of 66% was observed for the mixtures. The mixture cultivation of cover crops compensates the disadvantages of a single species [35]. The commonly cultivated cover crops and mixtures provide a large variability concerning the genera, physiology, morphology and the resulting weed suppressive ability for each species. Every single species in a mixture has individual properties to suppress weeds (with a broad target weed spectrum) and to adapt to varying biotic and abiotic field conditions. This can be followed by a higher flexibility and adaptability against stresses with an increased dry matter production and weed suppression compared to monocultures [28,35,67]. Beside higher biomass production, cover crop mixtures provide many ecological services compared to mono cultivated cover crops, e.g. higher biodiversity and erosion control [3,4]. For example, the cultivation of rye (S. cereale) and a legume, as summer vetch (V. sativa), in a mixture is able to produce higher amounts of biomass and N accumulation compared to a mono cultivation. This effect is based on ecological interaction, which allows an earlier germination and growth of rye and the ability of the vetch to climb on rye plants combined with atmospheric nitrogen accumulation [41,68–70]. The combination of these specific plant species properties provide a more rapid canopy closure with early weed suppressive effects [35,71,72]. Moreover, the low C:N ratio of this specific mixture prevents nitrogen immobilization with improved decomposition and nitrogen release rate for the following crop [35,70,73]. However, the influence of cover crop cultivation on the weed seed bank in autumn needs to be investigated. An increased biodiversity can promote the impact of seed predators, while their efficacy needs to be tested compared to herbicide application and mechanical approaches as stubble cultivation or false seed bed preparation.

No correlation was found between cover crop biomass and weed density which could be explained by the important role of biochemical effects on the overall weed suppression. Additionally, the large heterogeneity in the occurrence of weeds within the field may lead to insignificant correlations between these parameters [74].

The sowing date of the cover crops determines their germination, development speed, biomass accumulation and weed suppression during autumn and winter. In general, a cover crop should be sown shortly after the main crop harvest to avoid soil water losses due to evaporation and to confer a competition advantage to the cover crops against the weeds. The light interception by cover crops is negatively correlated with weed biomass which can be improved by an early and high light interception over short time compared to the whole season [75]. Therefore, a cover crop could be established as living mulch, sown shortly before the main crop harvest, to pass the seedling stage quickly and to generate a growth advantage against the weeds. After the main crop harvest, the increased amount of light promotes a rapid canopy closure with early, strong light interception to

suppress weeds efficiently. In chapter 2, the experimental years showed contrary results, which was probably because of soil water deficiencies. Further experiments should aim at the impact of available soil water and temperature at different cover crop sowing dates to induce optimum cover crop germination and development. A screening of different cover crops on drought tolerance could be performed to identify cover crops which were more suitable to unfavorable growth conditions. Based on this, specific cover crops could be selected for consistent germination and biomass production under varying field conditions in different years at early sowing dates. A further advantage of the early sowing would be the reduction of high workload peaks during and after main crop harvest while reducing the risk of delayed cover crop sowing with insufficient biomass production and weed control.

8.2 Inhibitory effects of cover crop mulches on weeds

Cover crops can produce high amounts of plant residues on the soil surface which can influence weed germination and growth in spring by reducing light transmittance, changing the microclimate, releasing allelochemicals and by building a physical barrier [3,35,44,71]. In chapter 5 and 6, the weed density was decreased by up to 83-97% across all locations and compared to the untreated control due to cover crop mulch. Furthermore, cover crop mulch originating from cover crop mixtures (-56%) tended to suppress weeds more effectively compared to monocultures (-31%). Significant weed reductions prior to sowing the main crop can lead to lower herbicide input accompanied by decreased environmental risks [4]. Nevertheless, the weed control is incomplete in many cases. The emerged weeds, which were not suppressed by cover crop mulch, were often sufficient to compete with the main crop and to generate significant yield reductions if no herbicides were applied [10]. While investigating the overall weed suppression by cover crop mulches, little information was available in literature regarding the changes in weed composition variation under different mulches [3]. In particular, large-seeded weeds, as Abutilon theophrasti Medicus, seemed to be less affected in germination by mulches compared to small-seed weeds as Chenopodium album [10,76]. The high impact of biochemical effects and physical parameters, e.g. bulk density, of cover crop mulch may be reasons for the observed insignificant correlation between cover crop mulch biomass and weed density. Therefore, the mulch biomass cannot be seen as the only indicator for weed control efficacy. Further parameters as the release of biochemical compounds, the mulch bulk density or the C:N ratio of the mulch should be implemented in future investigations on the weed suppressive ability of cover crop mulch. The release of allelochemicals by cover crop mulch can reduce weed germination and growth, whereby the suppressing effects are non-selective. Therefore, further experiments should aim on the release rate and persistence of allelochemicals in the soil to evaluate possible inhibitory effects on the following main crop.

8.3 Weed control with living mulches

Living mulches were shown to provide many benefits for an agricultural system by recycling nutrients, improving soil structure and suppressing weeds and pests [4,77,78]. They were used in many crops as vegetables, maize, cereals and oilseed rape [48–50,52,79–82]. The integration of living mulches in sugar beet crops is hardly performed due to the low competitiveness and slow development of sugar beet plants [51,83]. Interspecific competition for natural resources with quantitative and qualitative yield losses can limit this system [81]. In chapter 7, living mulches suppressed weeds by up to 71% and led to herbicide reductions of 65% in combination with prior hoeing and band-spraying compared to overall boom spraying. The cultivation of the living mulches 30 days after sugar beet sowing resulted in insignificant changes of white sugar yield and qualitative parameters compared to the overall boom spraying. An optimum sowing date, adequate living mulch species and cultivars, growth regulation and fertilization strategies in sugar beet crops need to be researched in future to enhance the acceptance of this cultivation method for farmers and to substantially reduce herbicide inputs in the long term.

8.4 Separation of biochemical and competitive effects by cover crops on specific weed species

Laboratory and greenhouse experiments were conducted to investigate the contribution of biochemical interaction between cover crops and weeds. During the last decades, several cover crop species were investigated for the active release of allelopathic compounds. Potential allelochemicals in the cover crops were isolated and identified as inhibitory compounds on several weed species [11,22–24,29]. Allelochemicals are naturally produced herbicides by plants with a relatively low phytotoxic activity compared to herbicides. The high efficacy of these compounds is obtained through the constant release into the environment with varying concentrations over long periods [84].

Beside other interference effects, allelopathy is an attractive explanation for observed weed suppression in many cover crop experiments, but the differentiation between all plant growth-influencing effects is difficult and diminishes the acceptance of this phenomenon in the agricultural science [59,85,86]. In plant research, nearly no other research field caused as much controversy as the studies on allelopathy [87,88].

8.4.1 Germinations tests with aqueous cover crop extracts

In chapters 3 and 5, different aqueous cover crop extracts were tested in germination tests to evaluate the biochemical inhibition by phytotoxic substances on germination, mean germination time and root length of specific weeds. Weeds are wild species and are genetically more heterogeneous compared to our crops and are characterized by a non-uniform germination [89]. Based on that, we selected representative weed species, which are naturally occurring in Germany with germination rates between 60 and 80%. This allows the detection of inhibitory and stimulatory effects combined with a low statistical variability. Monocotyledonous weeds were omitted due to possible autotoxic effects, which could affect germination [89,90].

The cover crop extracts inhibited weed germination and root growth significantly. Further, weed density within the field was correlated with the germination rate and root length of specific weeds from germination tests. This elucidates the important role of biochemical effects on the overall weed suppression within the field.

While using this experimental approach, the differentiation between allelopathic or phytotoxic effects on specific weeds constitutes a major challenge. In principle, phytotoxins which were isolated from a plant cannot be considered as allelochemicals *per se* without the knowledge about their role in plant communication or the environment [91]. Regardless of the initial purpose in the plant many substances were phytotoxic [23]. Based on that, germination tests with cover crop extracts were an important component in preliminary allelopathy studies to differentiate cover crops in their biochemical, suppressive activity on weeds [92].

Nevertheless, while testing cover crop extracts under laboratory conditions, a broad range of effects is eliminated due to the strong isolation of environmental factors. On the other hand, a field experiment would present a wide variability in response to the treatments, which necessitates the preliminary evaluation under isolated mechanisms with diverse model weeds [89]. However, chemical and biotic soil factors, the bioavailability and stability of allelochemicals in the soil as well as their uptake and response of the target plant are some of many factors which cannot be taken into account [93]. Moreover, preexisting compounds in the soil as methionine, glucose or nitrate and microbial communities were able to inhibit or promote the effects of allelochemicals in the soil [94,95].

Due to the complexity of the allelopathy phenomena, results from laboratory experiments are difficult to be transferred directly into natural field conditions [96]. However, we aimed to separate the biochemical weed suppressive effects of the cover crops from the overall weed suppression. Therefore, the relationship between the toxicity of cover crop extracts in germination tests and the weed density within the field was investigated. We observed a strong correlation between the mentioned parameters which allowed us to conclude, that biochemical effects are significantly involved on the overall weed suppression by cover crops in autumn and spring.

The inhibitory effects of the cover crop extracts varied along the different cover crops and between the investigated weeds. The highest weed suppressive effects were observed by *L. usitatissimum*, *H. annuus* and *T. subterraneum* which could

be explained by the possible release of biochemical substances during extraction of the cover crops. The cover crop F. esculentum showed a higher inhibition of weed growth in experiments with cover crop mulch incorporated in soil compared to the effects in germination tests. This indicates that some biochemical substances in the cover crop need to be transformed by microorganisms in the soil to become more active phytotoxic substances [27]. The comparison of the weed suppressive ability of *L. usitatissimum* in chapters 4 and 5 reveals that this specific cover crop shows no active release of allelochemicals during growth because no significant biochemical effects were measured in chapter 4. However, high weed suppressive effects were observed in experiments with cover crop extracts and mulch.

The experiments emphasized the species-specific activity of biochemical substances induced by cover crops. This could be due to differences in seed morphology and physiology or the ability of some weeds to detoxificate allelochemicals [28,65,97]. These observations may be used to achieve selective effects by allelochemicals to avoid inhibitory effects on the main crop.

8.4.2 Active carbon as adsorbent for allelochemicals

In chapter 4, active carbon in the soil was used as adsorbent for phytotoxic biochemical compounds released by different cover crops. The use of active carbon to reduce biochemical effects in the soil represents a suitable approach to evaluate the contribution of biochemical effects in the overall weed suppression [54,85,98–100]. Only a few experimental approaches have been conducted to exclude competitive effects as light, space, water and nutrients of crops and cover crops on weeds from biochemical interactions [64,101,102].

The active carbon in the soil bound chemical compounds like potential allelochemicals which provided a competitive disadvantage for the cover crops. Consequently, the biomass of some weeds was significantly increased under particular cover crops.

A correlation was calculated for the parameters cover crop and weed biomass for treatments *F. esculentum*, *A. strigosa*, *L. usitatissimum* and *H. annuus* cultivated in soil without active carbon (Soil-N) and for treatments *R. sativus*, *F. esculentum*,

L. usitatissimum in soil amended with active carbon (Soil-AC). In average, the correlation between cover crop and weed biomass was higher in Soil-N compared to Soil-AC. Under favorable growing conditions, an increasing amount of aboveground biomass of plants can be seen as an indicator for increasing amounts of root biomass and allelochemicals per plant [103–105]. Based on this, cover crops with a higher above-ground biomass will be more competitive against weeds due to higher competitive and biochemical effects, which could be shown in this experiment. If one factor for the competitive advantage of allelopathic cover crops is excluded, as biochemical effects due to the addition of active carbon in the soil, the influence of the cover crop biomass on weed biomass will decrease. If a specific cover crop lacks the ability to exert biochemical effects on the overall weed suppression, there will be no shift of competition between cover crop and weed. Consequently, the correlation between cover crop and weed biomass will not change, which was the case for cover crops L. usitatissimum and G. abyssinica in this study. Further studies, based on the results in chapter 4 and 5, should aim on the composition of cover crop mixtures with optimum morphological und physiological traits to suppress specific weed species or communities. Beside the weed suppression during cover crop growth, attention needs to be paid in cover crop selection regarding the weed suppressive ability of the mulch in spring due to different release ways of biochemical substances as observed for L. usitatissimum and F. esuclentum in chapters 4 and 5.

Moreover, the allelopathic effects can be species-specific and can affect the weed coexistence and the weed community composition [55,85,106]. This could be shown for the weed *S. media* as the most sensitive species to biochemical stress, which could also be observed in chapters 4 and 5.

However, active carbon can give inherent problems concerning the meaningfulness of the results. Lau et al. suggested, that the addition of active carbon in the potting soil may influence plant growth [59]. They found an increased nitrogen mineralization, if active carbon is present in the soil, which could explain a higher weed biomass in Soil-AC [59]. Furthermore, organic matter in the potting soil with allelopathic properties or changes in the soil microbial communities may affect the growth of the test plants [59,107]. Results in other publications dealing with minimizing allelopathic effects by the

incorporation of active carbon reveal that the growth effects can be inconsistent and can vary across the experimental conditions. For example, active carbon stimulated the growth of *Centaurea stoebe* (L.) ssp. *micranthos* in one study [59], but showed strong negative effects on *C. strobe micranthos* in another study [108]. Therefore, the disparity in the results in allelopathy studies highlights the importance of highly detailed methodological investigations in this research area to create standardized test systems in the future.

8.4.3 Further bioassays to test on allelopathy

The future research in allelopathy should aim at simple and standardized preliminary experiments to support or to refute, if a specific plant species is allelopathic or not [84]. The establishment of conditions in which plant growth, exudation and movement of the allelochemicals are as realistic as possible demonstrates a major challenge [85]. With this knowledge, different cover crop species and cultivars can be selected and might inhibit weed germination and growth more sustained which could generate noteworthy herbicide reductions before and in the following crop in spring and summer. To prove allelopathy, the allelopathic substances must be identified and characterized and could be further investigated as potential herbicidal compounds with new modes of action. In this section, continued studies in alleopathy with bioassays in the future were suggested.

1. A simple method to support allelopathy of a cover crop would be the cultivation of the donor (cover crop) and receiver plant (weed) side by side on agar. The exudated allelochemicals by the donor will diffundate through the medium and gradient effects on the root length of the receiver could be observed as indicator for the presence of allelopathic compounds [84,109,110].

2. An advanced approach of the first mentioned method can be the side by side cultivation of donor and receiver plant with a semipermeable root barrier. Consequently, this system would avoid direct root contact, while allelochemicals are able to move through the barrier [111,112]. On the other hand, a non-permeable membrane can be used to eliminate resource competition and the

movement of the allelochemicals between the plants [99]. Moreover, an opaque above-ground barrier would additionally exclude light competition [84,113].

3. The cultivation of the receiver plant in soil in which the donor plant was grown previously would exclude the resource competition. If potential allelochemicals were released into the soil during the growth of the donor, germination and/or growth reductions should be visible. In this experiment, a control treatment with a non-allelopathic cultivar of the same donor plant species should be created, as shown with rice cultivars in the study of Rimano and Duke [84,114]. Attention needs to be payed on the receiver plant density due to density-dependent effects of the allelochemicals [115]. This approach would enable us to identify biochemical inhibition levels along different cover crop cultivars. Moreover, the allelochemicals in the soil should be identified.

4. A further option is to cultivate the receiver plant in soil amended or irrigated with plant or soil leachates of the donor plant [116–118]. Due to the low half-life period of many allelochemicals, the draining water from donor plant pots could be transported directly to the donor plant pots [84]. Consequently, the leachates and root exudates would reach the receiver plant promptly.

5. The inhibitory effects on weeds occurring after the incorporation of cover crop mulch in soil in greenhouse experiments can differ strongly from observed effects within the field. The addition of active carbon in the pot medium, as adsorbent for secondary metabolites originating from the mulch, crossed with the exposure to allelopathic plant material, e.g. mulch, followed by enhanced weed growth can be interpreted as an indication for the presence of allelochemicals in the soil [58,99,109,119].

6. To include the preexisting environmental effects within the field, active carbon can be added in the field soil. After the incorporation, cover crops should be sown together with defined amounts of natural occurring weeds, while the preexisting weed flora should be eliminated to form a uniform weed infestation and to avoid a high variability in the results.

All of these methods would support the indication of allelopathic effects induced by the donor on the receiver plant. Nevertheless, many parameters as sampling time of plant biomass or compounds, climatic and soil conditions, irradiation, plant cultivar, fertilization or the type of charcoal can alter the allelopathic effects, which complicates the comparability of the experiments and results. However, the following identification, characterization and exploration of the mode-of-action of the released allelochemicals is needed to understand their physiological and ecological functions [85].

Chapter IX

Summary

Zusammenfassung

9 Summary

Weed control constitutes a major challenge in the worldwide crop production. Beside chemical and mechanical weed control strategies, cover cropping provides an effective way of biological weed suppression. Five different field experiments were conducted at six locations from 2014-2016 to evaluate the weed control efficacy of different cover crops in mono and mixed cultivation combined with different fertilization strategies and sowing dates. Furthermore weed suppressing effects of cover crop mulches in spring and of living mulches in summer were investigated. Potential effects on sugar beet emergence, quality and quantity were also assessed. In three laboratory and two greenhouse experiments from 2015-2017, the proportional contribution of competitive and biochemical effects on the overall weed suppression and the identification of varying susceptibilities of different weeds against biochemical stresses were at the center of research.

In field experiments, the weed suppressive effects of cover crops and living mulches in mono and mixed cultivation were tested. The experiments emphasized the importance of cover crop and living mulch mixtures compared to mono cropping due to a higher flexibility to biotic and abiotic stresses. This was followed by a more constant biomass production and more effective weed suppression. Moreover, the observed weed control was a result of competitive and biochemical effects, induced by cover crops. These were later on analyzed for active weed growth suppressing compounds.

Altering cover crop sowing date and fertilization to optimize the weed control resulted in significant changes of cover crop and weed biomass. Early cover crop sowing five or three weeks before winter wheat harvest increased the weed control efficacy in one year, significantly. Due to contrary results over the two experimental years, we suggest that the cover crop biomass and consequently the weed suppressive ability depends on sufficient soil water for rapid cover crop germination and growth.

The use of cover crop mulch in sugar beet crops provided a weed suppression of up to 83%. Especially mulch derived from cover crop mixtures reduced the weed

density (56%) more effectively compared to mono cultivated cover crops (31%). The inclusion of cover crops, mulches and living mulches can lead to significant herbicide reductions in the main crop. However supplementary mechanical or chemical weed control strategies are still necessary, especially in crops with a low competitive ability like sugar beets. Nevertheless, novel mechanical weed control approaches and adequate herbicide application techniques, as band-spraying, can reduce the herbicide input in the long-term.

Germination tests with aqueous cover crop extracts were conducted on weed seeds to evaluate differences in the inhibition of germination and root growth. Furthermore, different sensitivities of the weeds against the different cover crop extracts were revealed. Some cover crops as *S. alba, F. esculentum, H. annuus, T. subterraneum* and *L. usitatissimum* showed the most effective weed suppression. Moreover, the weed *M. chamomilla* showed the highest susceptibility against biochemical stresses in the germination tests.

A strong positive correlation between the weed suppressive effects by the extracts and the field weed suppression was found. This indicated that biochemical effects play also an important role on the overall weed suppression in the field. To estimate the proportions of competitive and biochemical effects on the overall weed suppression by cover crops, greenhouse experiments with active carbon supplemented soil were conducted. These experiments revealed that biochemical effects, by the presence of active carbon in the soil, shifted the balance of competition between cover crops and weeds. In the course of the experiments, we also found species-specific effects on the donor as well as on the receiver side.

The results of this thesis demonstrate the diverse use of cover crops, their mulches and living mulches in agricultural systems. This work aims on the optimization of biological weed control strategies and indicates approaches for future research. It is for example not yet clear how cover crops suppress specific weeds and if it is possible to design combinations of specific cover crops for the suppression of individual weed communities. Additionally, these results help to reduce long-term herbicide inputs in agricultural systems.

9.1 Zusammenfassung

Die Unkrautkontrolle stellt eine der größten Herausforderungen in der heutigen Pflanzenproduktion dar. Eine Eingliederung von Zwischenfrüchten und Untersaaten in die Fruchtfolge ist, neben chemischen und mechanischen Maßnahmen, eine effektive Möglichkeit zur biologischen Bekämpfung von Unkräutern.

In fünf verschiedenen Feldversuchen von 2014-2016 wurde die unkrautunterdrückende Wirkung von Zwischenfruchtreinsaaten und -mischungen in Kombination mit verschiedenen Anbauverfahren an sechs Standorten erforscht. Zudem wurden unkrautreduzierende Effekte durch den Einsatz von Zwischenfruchtmulch im Frühjahr und Untersaaten im Sommer im Hinblick auf Keimung, Wachstum, Qualität und Ertrag von Zuckerrüben untersucht. In drei Labor- und zwei Gewächshausversuchen von 2015-2017 sollte die Frage von den Anteilen von kompetitiven und biochemischen Effekten an der gesamten Unkrautunterdrückung beantwortet werden. Gleichzeitig sollten so auch Unterschiede im Wachstum einzelner Unkrautspezies aufgrund biochemischer Effekte von Zwischenfrüchten identifiziert werden.

Die Ergebnisse der Feldversuche deckten eine höhere Effektivität von Zwischenfruchtmischungen hinsichtlich der Unkrautkontrolle im Vergleich zu Zwischenfruchtreinsaaten auf. Diese beruhten auf der höheren Flexibilität der Zwischenfruchtmischungen gegenüber biotischen und abiotischen Stressoren, gefolgt von einer konstanteren und höheren Biomasse, die in diesen Versuchen zu einer effektiveren Unkrautunterdrückung führten. Darüber hinaus konnten diverse allelopathische Substanzen in den Zwischenfrüchten nachgewiesen werden, was auf eine Unkrautunterdrückung durch sowohl kompetitive als auch biochemische Effekte hinwies. Eine Veränderung des Aussaattermins und der Düngung der Zwischenfrüchte zur Optimierung der Unkrautkontrolle zeigte signifikante Effekte auf die Biomasse von Unkräutern und Zwischenfrüchten. Eine frühe Aussaat der Zwischenfrucht fünf und drei Wochen vor der Winterweizenernte erhöhte die Unkrautkontrolle signifikant in einem Jahr. Vermutlich hängen die der Zwischenfrüchte und die darauf Biomasseproduktion basierende

Unkrautunterdrückung von ausreichend pflanzenverfügbarem Bodenwasser für eine zügige Keimung und schnelles Wachstum ab.

Zwischenfruchtmulch auf der Bodenoberfläche im Frühjahr unterdrückte bis zu 83% der Unkräuter. Insbesondere der Mulch aus Zwischenfruchtmischungen konnte die Unkrautdichte durchschnittlich effektiver (56%) als Mulch aus Zwischenfruchtreinsaaten unterdrücken (31%). Weiterhin konnten durch den Zwischenfrüchten. Zwischenfruchtmulch Einsatz von und Untersaaten signifikante Herbizideinsparungen in der Hauptkultur erzielt werden. Durch Verfahren zur Unkrautbekämpfung wird meist nur biologische eine unvollständige Unkrautunterdrückung erreicht. Daher sind chemische oder Verfahren mechanische weiterhin notwendig, insbesondere bei konkurrenzschwachen Kulturen wie der Zuckerrübe.

In Laborversuchen wurden Keimtests mit wässrigen Zwischenfruchtextrakten an einzelnen Unkrautspezies durchgeführt. Anhand der Hemmung von Keimung und Wurzellänge wurden Unterschiede zwischen verschiedenen Zwischenfruchtextrakten identifiziert. Die Extrakte der Zwischenfrüchte S. alba, F. esculentum, H. annuus, T. subterraneum und L. usitatissimum zeigten die effektivste wachstumshemmende Wirkung. Das Unkraut M. chamomilla wies eine erhöhte Empfindlichkeit gegenüber den biochemischen Effekten dieser Zwischenfrüchte im Vergleich zu den anderen getesteten Unkräutern auf. Die wachstumshemmenden Effekte durch die Zwischenfruchtextrakte wurden mit der Unkrautunterdrückung aus Feldversuchen korreliert, wobei sich ein starker positiver Zusammenhang zeigte. Dies deutete auf einen großen Anteil an biochemischen Effekten an der gesamten Unkrautunterdrückung im Feld hin. Um die Anteile an kompetitiven und biochemischen Effekten von Zwischenfrüchten auf Unkräuter zu evaluieren, wurden Gewächshausversuche mit Aktivkohle versetztem Substrat durchgeführt. Diese Versuche zeigten, dass sich die unkrautunterdrückende Wirkung aufgrund von Konkurrenz durch die Zwischenfrucht in der Anwesenheit von Aktivkohle veränderte. Zudem konnten spezifische Effekte einzelner Zwischenfrüchte auf einzelne Unkrautspezies beobachtet werden.

Die Ergebnisse dieser Arbeit zeigen die diversen Vorteile von Untersaaten und Zwischenfrüchten sowie deren Mulch in landwirtschaftlichen Systemen. Diese Arbeit zielt auf eine Optimierung von biologischen Unkrautbekämpfungsmaßnahmen ab und zeigt neue Ansätze für zukünftige Forschung im Bereich der Zwischenfrucht-Unkraut Interaktion. Zudem tragen diese Ergebnisse dazu bei, einen weiteren Schritt zur langfristigen Reduzierung von Herbizideinträgen in der Landwirtschaft zu machen. **General References**

General References

1. Swanton, C. J.; Murphy, S. D. Weed Science Society of America Weed Science beyond the Weeds : The Role of Integrated Weed Management (IWM) in Agroecosystem Health Weed Science Beyond the Weeds : The Role of Integrated Weed Management (IWM) in Agroecosystem Health '. *Weed Sci* **2010**, *44*, 437–445.

2. Swanton, C.; Weise, S. Integrated weed management: the rationale and approach. *Weed Technol* **1991**, *5*, 657–663.

3. Campiglia, E.; Radicetti, E.; Mancinelli, R. Cover crops and mulches influence weed management and weed flora composition in strip-tilled tomato (Solanum lycopersicum). *Weed Res* **2015**, *55*, 416–425.

4. Hartwig, N. L.; Ammon, H. U. Cover crops and living mulches. *Weed Sci* **2002**, *50*, 688–699.

5. Weinert, T. L.; Pan, W. L.; Moneymaker, M. R.; Santo, G. S.; Stevens, R. G. Nitrogen recycling by nonleguminous winter cover crops to reduce leaching in potato rotations. *Agron J* **2002**, *94*, 365–372.

6. Danso, S. K. A.; Curbelo, S.; labandera, C.; pastorini, D. Herbage yield and nitrogen-fixation in a triple-species mixed sward of white clover, lotus and fescue. *Soil Biol Biochem* **1991**, *23*, 65–70.

7. Ding, G.; Liu, X.; Herbert, S.; Novak, J.; Amarasiriwardena, D.; Xing, B. Effect of cover crop management on soil organic matter. *Geoderma* **2006**, *130*, 229–239.

8. Wilson, G. F.; Lal, R.; Okigbo, B. N. Effects of cover crops on soil structure and on yield of subsequent arable crops grown under strip tillage on an eroded alfisol. *Soil Tillage Res* **1982**, *2*, 233–250.

9. Mendes, I.; Bandick, A.; Dick, R. Microbial biomass and activities in soil aggregates affected by winter cover crops. *Soil Sci Soc* **1999**, 873–881.

10. Teasdale, J. R. Contribution of cover crops to weed management in sustainable agricultural systems. *J Prod Agric* **1996**, *9*, 475–479.

11. Jabran, K.; Mahajan, G.; Sardana, V.; Chauhan, B. S. Allelopathy for weed control in agricultural systems. *Crop Prot* **2015**, *72*, 57–65.

12. Lawley, Y. E.; Weil, R. R.; Teasdale, J. R. Forage radish cover crop suppresses winter annual weeds in fall and before corn planting. *Agron J* **2011**, *103*, 137–144.

13. Haramoto, E. R.; Gallandt, E. R. Brassica cover cropping: I. Effects on weed and crop establishment. *Weed Sci* **2005**, *53*, 695–701.

14. Stivers-Young, L. Growth, nitrogen accumulation, and weed suppression by fall cover crops following early harvest of vegetables. *HortScience* **1998**, *33*, 60–63.

15. Teasdale, J. R.; Hatfield, J. L.; Buhler, D. D.; Stewart, B. A. Cover crops, smother plants, and weed management. *Integr weed soil Manag* **1998**, 247–270.

16. Lawley, Y. E.; Teasdale, J. R.; Weil, R. R. The mechanism for weed suppression by a forage radish cover crop. *Agron J* **2012**, *104*, 205–214.

17. Creamer, N. G.; Bennett, M. a.; Stinner, B. R.; Cardina, J.; Regnier, E. E. Mechanisms of weed suppression in cover crop-based production systems. *HortScience* **1996**, *31*, 410–413.

18. den Hollander, N. G. Growth characteristics of several clover species and their suitability for weed suppression in a mixed cropping design, 2012.

19. Rueda-Ayala, V.; Jaeck, O.; Gerhards, R. Investigation of biochemical and competitive effects of cover crops on crops and weeds. *Crop Prot* **2015**, *71*, 79–87.

20. Kruidhof, H.; Bastiaans, L.; Kropff, M. Cover crop-based ecological weed management : exploration and optimization; 2008.

21. Reigosa, M.; Pedrol, N.; González, L. Allelopathy - A Physiological Process with Ecological | Manuel J. Reigosa | Springer.

22. Kelton, J.; Price, A. J.; Mosjidis, J. Allelopathic weed suppression through the use of cover crops. *Weed Control* **2012**, 115–130.

23. Macías, F. A.; Molinillo, J. M. G.; Varela, R. M.; Galindo, J. C. G. Allelopathy - A natural alternative for weed control. *Pest Manag Sci* 2007, *63*, 327–348.

24. Singh, H. P.; Batish, D. R.; Kohli, R. K. Allelopathic Interactions and Allelochemicals: New Possibilities for Sustainable Weed Management. *CRC Crit Rev Plant Sci* **2003**, *22*, 239–311.

25. Soltys, D.; Krasuska, U.; Bogatek, R.; Gniazdowska, a. Allelochemicals as Bioherbicides - Present and Perspectives. *Herbic - Curr Res Case Stud Use* **2013**, 517–542.

26. Sanchez-Moreiras, a. M.; Weiss, O. a.; Reigosa-Roger, M. J. Allelopathic Evidence in the Poaceae. *Bot Rev* **2004**, *69*, 300–319.

27. Macías, F. A.; Oliveros-Bastidas, A.; Marín, D.; Chinchilla, N.; Castellano, D.; Molinillo, J. M. G. Evidence for an allelopathic interaction between rye and wild oats. *J Agric Food Chem* **2014**, *62*, 9450–9457.

28. Schulz, M.; Marocco, A.; Tabaglio, V.; Macias, F. A.; Molinillo, J. M. G. Benzoxazinoids in Rye Allelopathy - From Discovery to Application in Sustainable Weed Control and Organic Farming. *J Chem Ecol* 2013, *39*, 154–174.
29. Belz, R. G. Allelopathy in crop/weed interactions--an update. *Pest Manag Sci* 2007, *63*, 308–26.

30. Gianoli, E.; Niemeyer, H. M. Lack of costs of herbivory-induced defenses in a wild wheat: Integration of physiological and ecological approaches. *Oikos* **1997**, *80*, 269–275.

31. Jabeen, R.; Yamada, K.; Shigemori, H.; Hasegawa, T.; Hara, M.; Kuboi, T.; Hasegawa, K. Induction of ss-glucosidase activity in maize coleoptiles by blue light illumination. *J Plant Physiol* **2006**, *3*. *163*, 538–545.

32. Tinnin, R.; Muller, C. H. The Allelopathic Influence of Avena fatua : The Allelopathic Mechanism Author (s): Robert O. Tinnin and Cornelius H. Muller Published by : Torrey Botanical Society Stable URL : http://www.jstor.org/stable/2997070 of the Torrey Botanical Club The allel. **2017**, *99*, 287–292.

33. Gawronska, H.; Golisz, A. Allelopathy and biotic stresses. In *Allelopathy: A Physiological Process with Ecological Implications*; 2006; pp. 211–227.

34. Ridenour, W. M. W. W. M.; Callaway, R. M. R. The relative importance of allelopathy in interference: The effects of an invasive weed on a native bunchgrass. *Oecologia* **2001**, *126*, 444–450.

35. Teasdale, J.; Abdul-Baki, A. Comparison of mixtures vs. monocultures of cover crops for fresh-market tomato production with and without herbicide; 1998; Vol. 33, pp. 1163–1166.

36. Blanco-Canqui, H.; Shaver, T. M.; Lindquist, C. A. Cover Crops and Ecosystem Services: Insights from Studies in Temperate Soils. **2015**.

37. Chauhan, B. S.; Singh, R. G.; Mahajan, G. Ecology and management of weeds under conservation agriculture: A review. *Crop Prot* **2012**, *38*, 57–65.

38. Nichols, V.; Verhulst, N.; Cox, R.; Govaerts, B. Weed dynamics and conservation agriculture principles: A review. *F Crop Res* **2015**, *183*, 56–68.

39. Sheaffer, C. C.; Seguin, P. Forage Legumes for Sustainable Cropping Systems. *J Crop Prod* **2003**, *8*, 187.

40. Dahiya, R.; Ingwersen, J.; Streck, T. The effect of mulching and tillge on the water and temperature regimes of a loess soil:experimental findings and modeling. *Soil Tillage Res* **2007**.

41. Campiglia, E.; Caporali, F.; Radicetti, E.; R., M.; Mancinelli, R.; Radicetti, E.; Caporali, F. Effect of cover crops and mulches on weed control and nitrogen fertilization in tomato (Lycopersicon esculentum Mill.). *Crop Prot* **2010**, *29*, 354–363.

42. Ramakrishna, a.; Tam, H. M.; Wani, S. P.; Long, T. D. Effect of mulch on soil temperature, moisture, weed infestation and yield of groundnut in northern Vietnam. *F Crop Res* **2006**, *95*, 115–125.

43. Campiglia, E.; Radicetti, E.; Mancinelli, R. Weed control strategies and yield response in a pepper crop (Capsicum annuum L.) mulched with hairy vetch (Vicia villosa Roth.) and oat (Avena sativa L.) residues. *Crop Prot* **2012**, *33*, 65–73.

44. Teasdale, J. R.; Mohler, C. L. The quantitative relationship between weed emergence and the physical properties of mulches. *Weed Sci* **2000**, *48*, 385–392.

45. Moore, M. J.; Gillespie, T. J.; Swanton, C. J. Effect of Cover Crop Mulches on Weed Emergence, Weed Biomass, and Soybean (Glycine max) Development. *Weed Technol* **1994**, *8*, 512–518.

46. Kruidhof, H. M.; Bastiaans, L.; Kropff, M. J. Cover crop residue management for optimizing weed control. *Plant Soil* **2009**, *318*, 169–184.

47. Feil, B.; Liedgens, M. Pflanzenproduktion in lebenden Mulchen—eine Uebersicht. *Pflanzenbauwiss* **2001**.

48. Hiltbrunner, J.; Jeanneret, P.; Liedgens, M.; Stamp, P.; Streit, B. Response of weed communities to legume living mulches in winter wheat. *J Agron Crop Sci* **2007**, *193*, 93–102.

49. Hiltbrunner, J.; Liedgens, M.; Bloch, L.; Stamp, P.; Streit, B. Legume cover crops as living mulches for winter wheat: Components of biomass and the control of weeds. *Eur J Agron* **2007**, *26*, 21–29.

50. Masiunas, J. B. Journal of Vegetable Crop Production Production of Vegetables Using Cover Crop and Living Mulches—A Review. *J Veg Crop Prod* **1998**, *4*, 11–31.

51. Ahlers, G. Unkrautunterdrückung durch Untersaaten sowie Konkurrenzwirkung von Unkraut und Untersaaten auf die Zuckerrübe in Abhängigkeit der. **1986**.

52. Ilnicki, R. D. R.; Enache, A. J. A. Subterranean clover living mulch: an alternative method of weed control. *Agric Ecosyst Environ* **1992**, *40*, 249–264.

53. Rice, E. L. Allelopathy-An update. Bot Rev 1979, 45, 15–109.

54. Callaway, R. M.; Aschehoug, E. T. Invasive Plants Versus Their New and Old Neighbors: A Mechanism for Exotic Invasion. *Science* (80-) **2000**, 290, 521–523.

55. Prati, D.; Bossdorf, O. Allelopathic inhibition of germination by Alliaria petiolata (Brassicaceae). *Am J Bot* **2004**, *91*, 285–288.

56. Falquet, B.; Gfeller, A.; Pourcelot, M.; Tschuy, F.; Wirth, J. Weed Suppression by Common Buckwheat: A Review. *Environ Control Biol* **2015**, *53*, 1–6.

57. Macías, F.; Varela, R.; Torres, A.; Galindo, J. G.; Molinillo, J. G. Allelochemicals from sunflowers: chemistry, bioactivity and applications. *Chem Ecol Plants Allelopath Aquat Terr Ecosyst SE - 5* **2002**, 73–87.

58. Ridenour, W. M.; Callaway, R. M. The relative importance of allelopathy in interference: The effects of an invasive weed on a native bunchgrass. *Oecologia* **2001**, *126*, 444–450.

59. Lau, J. A. J.; Puliafico, K. K. P.; Kopshever, J. A. J.; Steltzer, H.; Jarvis, E. P.; Schwarzländer, M.; Strauss, S. Y.; Hufbauer, R. A. Inference of allelopathy is complicated by effects of activated carbon on plant growth. *New Phytol* **2008**, *178*, 412–423.

60. R Development Core, T. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, http://www.R-project.org/.

61. Bicksler, A. J.; Masiunas, J. B. Canada Thistle (Cirsium arvense) Suppression with Buckwheat or Sudangrass Cover Crops and Mowing. *Weed Technol* **2009**, *23*, 556–563.

62. Kumar, V.; Brainard, D. C.; Bellinder, R. R. Suppression of Powell Amaranth (Amaranthus powellii) by Buckwheat Residues: Role of Allelopathy. *Weed Sci* **2009**, *57*, 66–73.

63. Kalinova, J.; Vrchotova, N.; Triska, J. Exudation of allelopathic substances in buckwheat (Fagopyrum esculentum Moench). *J Agric Food Chem* **2007**, *55*, 6453–6459.

64. Falquet, B.; Roux, D.; Henriet, L.; Tschuy, F.; Wirth, J. Simple method to separate resource competition from allelopathic root interactions. *Allelopath J* **2014**, *34*, 227–240.

65. Haramoto, E. R.; Gallandt, E. R. Brassica cover cropping for weed management: A review. *Renew Agric Food Syst* **2004**, *19*, 187–198.

66. Sturm, D. J.; Kunz, C.; Gerhards, R. Inhibitory effects of cover crop mulch on germination and growth of Stellaria media (L.) Vill., Chenopodium album L. and Matricaria chamomilla L. *Crop Prot* **2016**, *90*, 125–131.

67. Altieri, M. A.; Lana, M. A.; Bittencourt, H. V.; Kieling, A. S.; Comin, J. J.; Lovato, P. E. Enhancing crop productivity via weed suppression in organic no-till cropping systems in Santa Catarina, Brazil. *J Sustain Agric* **2011**, *35*, 855–869.

68. Clark, A. J.; Decker, A. M.; Meisinger, J. J. Seeding rate and kill date effects on hairy vetch-cereal rye cover crop mixtures for corn production. *Agron J* **1994**, *86*, 1065–1070.

69. Clark, A. J.; Decker, A. M.; Meisinger, J. J.; McIntosh, M. S. Kill date of vetch, and a vetch-rye mixture: I. Cover crop and corn nitrogen. *Agron J* **1997**, *89*, 427–433.

70. Ranells, N. N.; Wagger, M. G. Nitrogen-15 recovery and release by rye and crimson clover cover crops. *Soil Sci Soc Am J* **1997**, *61*, 943–948.

71. Teasdale, J.; Mohler, C. Light transmittance, soil temperature, and soil moisture under residue of hairy vetch and rye. *Agron J* **1993**.

72. Jannink, J. L.; Liebman, M.; Merrick, L. C. Biomass production and nitrogen accumulation in pea, oat, and vetch green manure mixtures. *Agron J* **1996**, *88*, 231–240.

73. Clark, A.; Decker, A.; Meisinger, J. Kill date of vetch, rye, and a vetch-rye mixture: II. Soil moisture and corn yield. *Agronomy* **1997**.

74. Hodgdon, E. A.; Warren, N. D.; Smith, R. G.; Sideman, R. G. In-Season and carry-over effects of cover crops on productivity and weed suppression. *Agron J* **2016**, *108*, 1624–1635.

75. Kruidhof, H.; Bastiaans, L.; Kropff, M. Ecological weed management by cover cropping: effects on weed growth in autumn and weed establishment in spring. *Weed Res* **2008**, *48*, 492–502.

76. Putnam, A. R.; DeFrank, J. Use of phytotoxic plant residues for selective weed control. *Crop Prot* **1983**, *2*, 173–181.

77. Carof, M.; Tourdonnet, S.; Saulas, P.; Floch, D.; Roger-Estrade, J. Undersowing wheat with different living mulches in a no-till system. II. Competition for light and nitrogen. *Agron Sustain Dev* **2007**, *27*, 357–365.

78. Nakamoto, T.; Tsukamoto, M. Abundance and activity of soil organisms in fields of maize grown with a white clover living mulch. *Agric Ecosyst Environ* **2006**, *115*, 34–42.

79. Liedgens, M.; Frossard, E.; Richner, W. Interactions of maize and Italian ryegrass in a living mulch system:(2) Nitrogen and water dynamics. *Plant Soil* **2004**, *259*, 243–258.

80. White, J. G.; Scott, T. W. Effects of perennial forage-legume living mulches on no-till winter wheat and rye. *F Crop Res* **1991**, *28*, 135–148.

81. Masiunas, J. B. Production of Vegetables Using Cover Crop and Living Mulches — A Review. *J Veg Crop Prod* **1998**, *4*, 11–31.

82. Enache, A. J.; Ilnicki, R. D. Weed control by subterranean clover (Trifolium subterraneum) used as a living mulch. *Weed Technol* **1990**, *4*, 534–538.

83. Haberland, R. Effect of clover intercrops on weed infestation and yield of sugar beet. *Gesunde Pflanz* **1996**, *48*, 199–204.

84. Duke, S. O. Proving Allelopathy in Crop–Weed Interactions. *Weed Sci* **2015**, *63*, 121–132.

85. Inderjit; Callaway, R. M. Experimental designs for the study of allelopathy. *Plant Soil* **2003**, *256*, 1–11.

86. Hierro, J. L.; Callaway, R. M. Allelopathy and exotic plant invasion. *Plant Soil* 2003, *256*, 29–39.

87. Kruse, M.; Strandberg, M.; Strandberg, B. *Ecological Effects of Allelopathic Plants – a Review Ecological Effects of Allelopathic Plants – a Review*; 2000.

88. Romeo, J.; Weindenhamer, J. D. Bioassays for allelopathy in in terrestrial plants. *Methods Chem Ecol Vol 2 Bioassays methods* **1998**, *II*, 179–211.

89. Macías, F. A.; Castellano, D.; Molinillo, J. M. G. Search for a standard phytotoxic bioassay for allelochemicals. Selection of standard target species. *J Agric Food Chem* **2000**, *48*, 2512–2521.

90. Waller, G. R.; Krenzer, E. G.; McPherson, J. K.; McGown, S. R. Allelopathic compounds in soil from no tillagevs conventional tillage in wheat production. *Plant Soil* **1987**, *98*, 5–15.

91. Duke, S. O. Phytotchemical phytotoxins and hormesis - A commentary. *Dose-Response* **2011**, *9*, 76–78.

92. Leather, G. R.; Einhellig, F. A. Bioassays in the study of allelopathy. In *The Science of Allelopathy*; 1986; pp. 133–145.

93. Kruidhof, M.; Gallandt, E. R.; Haramoto, E. R.; Bastiaans, L. Selective weed suppression by cover crop residues: Effects of seed mass and timing of species' sensitivity. *Weed Res* **2011**, *51*, 177–186.

94. Inderjit; Nilsen, E. T. Bioassays and Field Studies for Allelopathy in Terrestrial Plants: Progress and Problems. *CRC Crit Rev Plant Sci* **2003**, *22*, 221–238.

95. Blum, U.; Gerig, T. M.; Worsham, A. D.; King, L. D. Modification of allelopathic effects of p-coumaric acid on morning-glory seedling biomass by glucose, methionine, and nitrate. *J Chem Ecol* **1993**, *19*, 2791–2811.

96. Inderjit; Weston, L. A. Are laboratory bioassays for allelopathy suitable for prediction of field responses? *J Chem Ecol* **2000**, *26*, 2111–2118.

97. Schulz, M.; Marocco, A.; ecology, V. T.-J. of chemical; 2012, undefined BOA detoxification of four summer weeds during germination and seedling growth. *Springer*.

98. Mahall, B. E.; Callaway, R. M. Root communication mechanisms and intercommunity distributions of two mojave desert shurbs. *Ecology* **1992**, *73*, 2145–2151.

99. Nilsson, M. C. Separation of allelopathy and resource competition by the boreal dwarf shrub Empetrum hermaphroditum Hagerup. *Oecologia* **1994**, *98*, 1–7.

100. Kulmatiski, A.; Beard, K. H. Activated carbon as a restoration tool: Potential for control of invasive plants in abandoned agricultural fields. *Restor Ecol* **2006**, *14*, 251–257.

101. Tschuy, F.; Gfeller, A.; Azevedo, R.; Khamisse, C.; Henriet, L.; Wirth, J. Weed suppression by cover crops: analyzing different factors. *Agrar Schweiz* **2014**, *5*, 292–299.

102. He, H. Bin; Wang, H. Bin; Fang, C. X.; Lin, Z. H.; Yu, Z. M.; Lin, W. X. Separation of allelopathy from resource competition using rice/barnyardgrass mixed-cultures. *PLoS One* **2012**, *7*.

103. Clossais-Besnard, N.; Larher, F. Physiological role of glucosinolates in {IBrassica napus}. Concentration and distribution pattern of glucosinolates among plants organs during a complete life cycle. *J Sci Food Agric* **1991**, *56*, 25–38.

104. Luo, W.; Jiang, Y.; Lü, X.; Wang, X.; Li, M. H.; Bai, E.; Han, X.; Xu, Z. Patterns of Plant Biomass Allocation in Temperate Grasslands across a 2500-km Transect in Northern China. *PLoS One* **2013**, *8*, 1–8.

105. Monk, C. Ecological importance of root/shoot ratios. *Bull Torrey Bot Club* **1966**.

106. Belz, R. G.; Reinhardt, C. F.; Foxcroft, L. C.; Hurle, K. Residue allelopathy in Parthenium hysterophorus L.-Does parthenin play a leading role? *Crop Prot* **2007**, *26*, 237–245.

107. Pietikäinen, J.; Kiikkilä, O.; Fritze, H. Charcoal as a habitat for microbes and its effect on the microbial community of the underlying humus. *Oikos* **2000**, *89*, 231–242.

108. Newingham, B. A.; Callaway, R. M. Shoot herbivory on the invasive plant, Centaurea maculosa, does not reduce its competitive effects on conspecifics effects and natives. *Oikos* **2014**, *114*, 397–406.

109. Bertin, C.; Harmon, R.; Akaogi, M.; Weidenhamer, J. D.; Weston, L. A. Assessment of the phytotoxic potential of m-tyrosine in laboratory soil bioassays. *J Chem Ecol* **2009**, *35*, 1288–1294.

110. Bertin, C.; Weston, L. A.; Huang, T.; Jander, G.; Owens, T.; Meinwald, J.; Schroeder, F. C. Grass roots chemistry: meta-Tyrosine, an herbicidal nonprotein amino acid. *Proc Natl Acad Sci* **2007**, *104*, 16964–16969.

111. Hilt, S.; Beutler, E.; Bauer, N. Comparison of Methods To Detect Allelopathic Effects of Submerged Macrophytes on Green Algae. *J Phycol* **2012**, *48*, 40–44.

112. Fujii, Y.; Pariasca, D.; Shibuya, T.; Yasuda, T.; Kahn, B.; Waller, G. R. Plant-box Method: A Specific Bioassay to Evaluate Allelopathy through Root Exudates. In *Allelopathy: New Concepts and Methodology*; 2007; pp. 39–56.

113. Wu, H.; Pratley, J.; Lemerle, D.; Haig, T. Laboratory screening for allelopathic potential of wheat accessions against annual ryegrass. *Aust J Agric Res* **2000**, *51*, 259–266.

114. Rimando, A.; Duke, S. Studies on rice allelochemicals; 2003.

115. Weidenhamer, J. D.; Hartnett, D. C.; Romeo, J. T. Density-Dependent Phytotoxicity: Distinguishing Resource Competition and Allelopathic Interference in Plants. *J Appl Ecol* **1989**, *26*, 613–624.

116. Hagan, D. L.; Jose, S.; Lin, C. H. Allelopathic Exudates of Cogongrass (Imperata cylindrica): Implications for the Performance of Native Pine Savanna Plant Species in the Southeastern US. *J Chem Ecol* **2013**, *39*, 312–322.

117. Chou, C.-H.; Muller, C. H. Allelopathic Mechanisms of Arctostaphylos glandulosa var. zacaensis. *Am Midl Nat* **1972**, 88, 324.

118. Dietz, H.; Steinlein, T.; Winterhalter, P.; Ullmann, I. Role of allelopathy as a possible factor associated with the rising dominance of Bunias orientalis L.(Brassicaceae) in some native plant assemblages. *J Chem Ecol* **1996**, *22*, 1797–1811.

119. Swain, D.; Paroha, S.; Singh, M.; Subudhi, H. N. Evaluations of allelopathic effect of Echinochloa colona weed on rice (Oryza sativa L. "Vandana"). *J Environ Biol* **2012**, *33*, 881–889.

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